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Nucleotide sequences from the terminal regions of fowl plague virus genome RNA

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The genome of influenza virus consists of eight discrete single-stranded RNA segments each of which codes for a unique polypeptide species (McGeoch et al. 1976; Inglis et al. 1977; Inglis & Almond, this symposium). These viral polypeptides are synthesized from mRNA molecules that are complementary to the viral RNA (vRNA) and the 3' ends of which lack sequences corresponding to the 5' terminus of vRNA (Hay et al. 1977). I have determined the nucleotide sequences of the 5' and 3' termini of the eight RNA segments of fowl plague virus (FPV), an avian influenza A strain, by recently developed methods for direct RNA sequencing. These methods involve radiolabelling of the RNA segments in vitro at either the 5' end or the 3' end followed by partial digestion of individual segments with specific endoribonucleases and analysis of the products by one-dimensional or two-dimensional polyacrylamide gel electrophoresis (Donis-Keller et al. 1977; Simoncsits et al. 1977; England & Uhlenbeck 1978; Lockard et al. 1978).

Table 1 shows the nucleotide sequence of the 5' terminal regions of the eight RNA segments of FPV. Each segment contains a common region of 13 nucleotides at the 5' end as has been reported previously by Skehel & Hay (1978), followed by a distinctive triplet sequence (underlined in table 1) which is unique in each segment, with the exception of segments 4 and 8 (the HA and NS genes respectively) which contain an identical triplet. These triplets are followed by a region of 5–7 uridine residues after which each segment contains a unique nucleotide sequence for the additional 40–60 nucleotides analysed. The 5' terminal region of vRNA, with the exception of the first 25–30 residues, corresponds to the 3' end of mRNA (Skehel & Hay 1978) and several protein termination codons can be identified (not shown); however, whether or not any of these termination codons are utilized *in vivo* remains to be determined.

The nucleotide sequences derived from the 3' termini of the RNA segments of FPV are shown in table 2 and reveal a common region of 12 nucleotides at the extreme 3' end of each segment. These sequences complement those reported by Skehel & Hay (1978) who found a common sequence of 12 nucleotides at the 5' end of *in-vitro* transcripts of vRNA. Beyond this common region each segment contains a unique sequence. It was first observed by Skehel & Hay (1978) that in any one segment a hexanucleotide comprising the distinctive triplet underlined in table 1 plus the adjacent three nucleotides of the common region (i.e. residues 11–16 of the 5' end) was exactly complementary to a hexanucleotide comprising the triplet underlined in table 2 plus the adjacent three nucleotides of the common region (i.e. residues 10–15 of the 3' end). Tables 1 and 2 indicate that this complementarity exists in segments 3, 5, 6, 7 and 8 but not in segment 4. Preliminary data (not shown) suggest that this complementarity also exists in segments 1 and 2.

The 5' terminal nucleotide sequence of influenza mRNA can be deduced by complementarity

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TABLE 1. 5' TERMINAL SEQUENCES OF FOWL PLAGUE VIRUS

	JUCCA	ACAGGACUCUGCUGUCCUGCGGA
50	UAUCAUUAAAUAAGCUGAAAGGAGAAAGUUCUUAUCUUUGCUCCA	JUACUCCAGCUCUAUGUUGACAAAUGACCAUCGUCAAAAUCCACACCACUCUGCUCUCCUCCCCAA
25	AGUAGAAACAAGGUGUUUUUUUUAUCAUUAAAUAAGC	AGUAGAAAGAAGGUAGUUUUUUUAGUGGAGGUGUAUG L
ment	8 AGUAGAAACAA	7 AGUAGAAAGAA

AGUAGAAACAAGGAGUUUUUUUGAACAAAQUAGUUGUCAAUGGUGAAUGGGAACUCAGCACCGUCUGG

AGUAGAAACAAGGGUAUUUUUUUUUUAAUUGUCAUACUCCUCUGCAUUGUCUCGGAAGAAAUAAGAUCCCUCAUUA 10

AGUAGAAACAAGGUACUUUUUGGACAGUAUGGAUAGCAAAUAGUAGCAUUGCCACAAACUAUUUCAGUGCAUG က

4

AGUAGAAAGAAGGCAUUUUUUQAUGAAGGACAAGCUAAAUUCAGUUUUUCUGCGGUCUGAGCUCUU

AGUAGAAACAAGGUGGUUUUUAAACAAUUGGACAUUAAUUGAUGGCCAUCCGAAUUCUUUGG

TABLE 2. 3' TERMINAL SEQUENCES OF FOWL PLAGUE VIRUS

segment	œ	7	9	ιĊ	4	က	63	T
50 25	AAAGAAAGCAGUCUACCUGAAAGCUUGACACAGUGUUGGAAUCCAUUAUGUUUUUGUCACCCUGCUUUUUGCU	CGGGACGACAGAGAAGGUAGGUUCGAAGCUCGGUUAGAAGACUCAUCUUAAAUAUCUACGUGCUUUUGGU	GACCAAUGGUUAUUAUUUUGUGAUUUGGAUUCAUUUUGAAGUCGUUUUUGGU	GUCCUGAGACCCAUGAVAUGGACCCACCCACUCAGUGACUCAUVAVAUACCCUGCUUUUGCU	GCAAGGGCGAAAACCAGGAUTUGAGUGUUCAUTUUGUAAGCCCUGCUUUUGCU	GGAUUGAAGCAUUGACGCACAAAUUCUUCCAUUUGGAUCAGUACCUGCUUUUGCU	CCUGCUULUGGU	CCACCAAAAA

FOWL PLAGUE VIRUS TERMINAL SEQUENCES

Table 3. 5' Terminal sequences of the MRNA of fowl plague virus

		Pro	C
\mathbf{Leu}	CCC	Val	CITC
	$\Omega\Omega\Omega$		
	UGC		
Asp	GAC	Len	
Val	GUA	Val	CIII
Gla	J CAG GUA	Tvr	TIAC
\mathbf{Phe}	DO	\mathbf{Thr}	V
Ser	AGC	Glv	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \
Ser	GUG UCA	Val	CIIII
Val	GUG	Glu	CA.C
Thr	ACU	Thr	ACC
Asn	AAC	Len	CITA
Ser	DCC	Len	CLII
Asp	GAU	Ser	AGI
25	AGCAAAAGCAGGGUGAGAAAAAACAUAAUG GAU		AGGAAAAGGAGGIJAGAIJIIJAAAGAIJG
	œ		7

Asn Pro Asn Gln Lys Ile Ile Thr Ile Gly AAU CGA AAU GAG AAA AUA AUA ACC AUU GGG Ser Gln Asp UCU CAG GAC

Glu Glu Phe Val Arg Gln Cys Phe Asn Pro GAA GAA UUU GUG CGU CAA UGC UUC AAU CC Asn Thr Gln Ile Leu Val Phe Ala Leu Ala AGGAAAAGGAGGGUUAGAAAAUG AAG ACU CAA AUG CUG GUU UUG GCG CUU GC Ala AGCAAAAGCAGGGUAUAUAAUCACUGACUGAGGGGGCGUCCAUAUCAUG GCG

AGGAAAAGCAGGUACUGAUCGAAAAUG

† Deduced by complementarity from the 3' terminal nucleotide sequence of genome RNA (table 2).

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AGCAAAAGCAGGAGUUCAAAAUG

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from the 3' end of vRNA and is shown in table 3. These data do not include any primer sequences found in vivo at the 5' end of mRNA and which are not complementary to vRNA (Bouloy et al. 1978; Krug et al., this symposium). The first and only AUG protein initiation codon located in each mRNA species is underlined in table 3 and is located 20-30 residues from the 5' end in all species except mRNA 5. Preliminary data (not shown) also locate the first AUG codon in mRNAs 1 and 2 in this region. In mRNA 5, the first AUG codon occurs at residues 46-48. It is of interest that the AUG codons in mRNAs 3, 4 and 6 are all located in the unique sequence 5'CAAAAUG-3'. A comparison of all known eukaryotic 5' mRNA sequences indicates that in nearly all cases the AUG closest to the 5' end is responsible for initiation and a model for ribosome binding and protein synthesis initiation incorporating this observation has been proposed (Kozak 1978). Thus although there is no direct evidence, by comparison with other eukaryotic mRNAs it is highly probable that these AUG codons identified in table 3 are utilized for protein synthesis initiation. The putative terminal amino acid sequences of the corresponding viral proteins can be deduced from the nucleotide sequences and are indicated in table 3. The predicted amino acid sequence for segments 4 and 6, which code for the haemagglutinin and neuraminidase glycoproteins respectively, contain a high proportion of hydrophobic amino acid residues. This is compatible with the proposed 'signal' sequence of amino acids which should be found in the extreme amino-terminus of membrane glycoproteins. Several termination codons can be identified (not shown), none of which are located in the same reading frame as the predicted amino acid sequence and thus do not contradict the proposed structures. More direct evidence either from complete nucleotide sequence analysis of the segments or from direct amino acid sequencing will be required to determine whether or not the mRNA species direct protein synthesis as shown in table 3.

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